## Andreas Teske Laboratory ICoMM DNA Extraction Protocol:

## Sample collection

Sediment samples were collected on-ship via APC (advanced piston coring) and frozen at -80°C (1). Samples were shipped on dry-ice to the University of North Carolina at Chapel Hill and were kept at -80°C until analysis. Sediment cores from Ocean Drilling Program (ODP) Leg 201, Site 1227, 1229 and 1231 were used in this study <sup>1</sup>. Sites 1227 and 1229 are on the Peru Margin

## **DNA** extraction

- 1. Aseptically scrape sediment surface.
- 2. Discard the first centimeter of sediment material.
- 3. Collect subsequent scrapings in a sterile container (petri dish), homogenize and weigh until there is 4 grams.
- 4. Prepare to use the MoBio UltraClean Soil DNA kit <sup>2</sup>(MoBio Laboratories, Inc., Carlsbad, CA, catalog number 12800-50) with the following modifications
- 5. Follow the manufacturer protocol:
- 6. To the 2ml Bead Solution tubes provided, add 0.5 gm of sample. Gently vortex to mix.
- 7. Add 60µl of Solution S1 and invert several times or vortex briefly.
- 8. Add 200µl of Solution IRS (Inhibitor Removal Solution), vortex briefly.
- 9. Place samples in a 65°C waterbath for 20 minutes.
- 10. Homogenize in a bead beater for 1 minute.
- 11. Centrifuge samples for 1 minute
- 12. Perform the remaining steps as per manufacturer protocol (step 8).
- 13. Pool elutions of extractions and use 1-10 microliters for PCR template
- 14. **\*\***Note, for these samples PCR was done using HiFi Platinum polymerase. Pfu polymerase did not amplify these samples well.

## **References:**

 D'Hondt SL, Jørgensen BB, Miller DJ, et al. (2003) Proceedings of the Ocean Drilling Program, Initial Reports, Leg 201 [CD-ROM]. Available from: Ocean Drilling Program, Texas A&M University, College Station TX 77845-9547, USA.
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